APARTS



Antwerp PAthology Research and Trial Service **INQUIRY & QUOTE REQUEST FORM**

Email: APARTS@uza.be

PROJECT INFORMATION

Project short title / Acronym				
Title				
Sponsor/ Funding institution				
MEC approval	obtained Ref:	pend	ing	N/A
Mandatory Biobank agreement (link to Biobank)	YES Ref:	NO		N/A
Principle Investigator (PI)				
Institution & department				
Purchase Order number / Project (budget) ID				
Contact person				
E-mail contact				
Project type	Academic		Commer	cial
Co-publication ^{\$}	YES		NO	
Nature of Request	Cost estimate fo project proposal	r	Approve	d project
Proposed delivery date of APARTS service (MM/YYYY)				
Number of cases				
Diseases type				
Sample Type	Tissue:	FFPE tissue	FF tissue	
	Fluids:	Urine	Blood	Plasma
	Other:			

^{\$} A discount may apply for the analysis fee of molecular biologist/pathologist in case of co-publication.

UA/UZA pathology group	No
member involved in the	Yes – Name:
project?	Type of involvement (e.g. copromotor):

TYPE OF ANALYSIS

- 1. Sample processing and staining (histology, immunohistochemistry and ISH)
- 2. Histopathologic evaluation
- 3. Digital Microscopy
- 4. Molecular analysis

1 Sample processing and staining

The principle investigator has obtained the appropriate consent, medical ethical commission (MEC) permission and proper use of the biomaterials.

Processing			
Fixation	formalin	ethanol	other:
Embedding	paraffin blocks	cell suspension in agar cell blocks	other:
	other:		
Number of blocks			

Please provide an excel list with sample names. This name will be printed on the cassette.

Cutting						
Sample type	paraffin blocks		frozen	tissue	TMA	blocks
Tissue type (i.e. organ)						
Total no. of blocks						
Number of sections	No. of blocks					
	Section thickness (μM)					
Specification per block	No of sections/block					
Method	RNAse-free		serial sec	ctions on	section	ons in cups
	cutting	gl	ass			
Planned analysis	Histochem		IHC or (F)ISH	D/ RI	NA-isolation
	Other, i.e		•			
Other remarks			•			

Histochemical stainings				
Staining	HE	PAP	PAS	other:
Number of slides				

Immunohistochemical st	Immunohistochemical stainings					
	IHC staining on Omnis (Dako/Agilent) or BenchMark Ultra (Ventana/Roche) Only for human FFPE samples					
Protocol	UZA PA*: see <u>list IHC UZA pathology department</u> optimisation/validation in consultation with researcher protocol (for Omnis/Benchmark) supplied by researcher					
Antibody	# AB provided by researcher*					
Control tissue	each run each slide not required (e.g. internal control present)					
Other remarks						
IHC technical quality will	be evaluated b	y patholog	gy dep	artment		

^{*} if protocol of UZA PA is used, then AB is provided by APARTS

In Situ Hybridisaition (IS	In Situ Hybridisaition (ISH)					
Material	FFPE tissue slides	Cytospins	Other, i.e			
	List of probes*: see <u>list</u>	FISH UZA pathology depai	rtment			
(F)ISH	Probe	#	Probe provided by researcher			
(F)ISH						
Tune of analysis	ANADUELCATION	DDEAK FU	CION DELETION			
Type of analysis	AMPLIFICATION	BREAK FUS	SION DELETION			
Number of nuclei for evaluation	20	50	Other:			
Representative						
microscopic image	YES NO					
needed						
CISH	EBER	Other:				

^{*} if protocol of UZA PA is used, then probe is provided by APARTS

2 Histopathologic evaluation

Microscopic evaluation						
	evaluate if representative tissue is present					
H&E	% relevant cells		ROI selection and annotation			
	other:					
	#	Scoring system	Scoring system			
ІНС		H-score: intensity & %	other scoring system:	descriptive		
Evaluation by a specific	No		•			
pathologist requested?	Yes – Name:					
Other remarks						

3 Digital Microscopy (WSI, Ultra-fast scanner, Philips, 40x)

Slides need to be clean without excess mounting media on the (edges of the) cover glass, mounting medium has to be dry and cover glass should NOT stick out. No marking on the slide allowed.

Please provide an external hard drive for image storage (0.5-1.5 Gb/image at 40x, depending on tissue area).

Scanning					
Number of slides					
Image format	Tiff	ISyntax	(
Required magnification:	40x	20x	10x	5x	
Back-up storage for 1 month by APARTS	YES	NO			

4 Molecular analysis

NA extraction			
D/RNA isolation of	# of cases:	Macrodissection requir	red? YES NO
FFPE sections	# of sections /case:		
cfDNA isolation	Material: plasma	urine CSF	other:
CIDINA ISOIALIOII	# of cases:	ml/case:	Elution volume:
Measurement of	Nanodrop	Qubit DNA BR	Qubit DNA HS
concentration	Qubit RNA BR	Qubit RNA HS	
Other remarks			

PCR			
	BRAF V600	NRAS+BRAF	KRAS
Idylla	NSCLC fusion	MSI	
	ctEGFR	ctKRAS	
	Other:		
Real-time PCR	HPV Xpert (cyto)	HPV sacace (cyto +	
	FFPE)		
	Other:		
	KRAS G12/13	KRAS G12C	BRAF V600
Digital droplet PCR	EGFR 5 hotspots	EGFR T790M	MYD88 L265P
Digital di opiet PCN	Other:		

Full range of available assays on Idylla platform and QX200 ddPCR platform: <u>biocartis website</u> en <u>biorad website</u>.

Nex	t Generation Sequencing (NGS)
	Oncomine Focus Assay (OFA) – DNA panel (SNV and small indels, 23 genes)
	Oncomine Focus Assay (OFA)— RNA panel for fusions (22 genes) + METex14 skipping & EGFR
	VIII
	Oncomine Comprehensive Assay (OCA) – DNA panel (143 genes)
	Oncomine Comprehensive Assay (OCA) – RNA panel (22 fusions)
	BRCA1/2 with Oncomine Research Assay (SNV and small indels)

Gynaecology targeted NGS panel (22 genes)
Glioma targeted NGS panel including TP53, IDH1/2, 1p19q deletion and EGFR amplification
Thyroid targeted NGS panel including POLE (15 genes)
TMB with Oncomine Tumor Mutation Load assay
CNA by sWGS
Oncomine PAN-CANCER cell-free assay (52 genes)
Other:

Detailed NGS panel information is available here: <u>labogids UZA pathologie - NGS</u>.

MGMT promoter hypermethylation by RT-PCR MLH1 promoter hypermethylation by RT-PCR Whole genome methylation by Infinium EPIC microarray Other: