**APARTS**

**A**ntwerp **PA**thology **R**esearch and **T**rial **S**ervice

INQUIRY & QUOTE REQUEST FORM

Email: aparts@uza.be

**PROJECT INFORMATION**

|  |  |  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- | --- | --- |
| Project short title / Acronym |  | | | | | | |
| Title |  | | | | | | |
| Sponsor/ Funding institution |  | | | | | | |
| MEC approval | obtained  Ref: … | | pending | | | N/A | |
| Mandatory Biobank agreement [*(link to Biobank)*](https://www.uza.be/research/biobank-antwerpen/stalen-aanvragen-enof-registreren) | YES  Ref: … | | NO | | | N/A | |
| Principle Investigator (PI) |  | | | | | | |
| Institution & department |  | | | | | | |
| Purchase Order number / Project (budget) ID |  | | | | | | |
| Contact person |  | | | | | | |
| E-mail contact |  | | | | | | |
| Project type | Academic | | | | Commercial | | |
| Co-publication$ | YES | | | | NO | | |
| Nature of Request | cost estimate for project proposal | | | | approved project | | |
| Proposed delivery date of APARTS service (MM/YYYY) |  | | | | | | |
| Number of cases |  | | | | | | |
| Diseases type |  | | | | | | |
| Sample Type | Tissue: | FFPE tissue | | FF tissue | | |  |
| Fluids: | Urine | | Blood | | | Plasma |
| Other: … | | | | | | |

*$ A discount may apply for the analysis fee of molecular biologist/pathologist in case of co-publication.*

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| --- | --- |
| UA/UZA pathology group member involved in the project? | No  Yes – Name: …  Type of involvement (e.g. copromotor): … |

**TYPE OF ANALYSIS**

1. **Sample processing and staining (histology, immunohistochemistry and ISH)**
2. **Histopathologic evaluation**
3. **Digital Microscopy**
4. **Molecular analysis**

**1 Sample processing and staining**

 *The principle investigator has obtained the appropriate consent, medical ethical commission (MEC) permission and proper use of the biomaterials.*

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| **Processing** | | | |
| **Fixation** | formalin | ethanol | other: … |
| **Embedding** | paraffin blocks | cell suspension in agar cell blocks | other: … |
| Other: … | | |
| **Number of blocks** |  | | |

Please provide an excel list with sample names. This name will be printed on the cassette.

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| **Cutting** | | | | | | | | |
| **Sample type** | paraffin blocks | | frozen tissue | | | | TMA blocks | |
| **Tissue type (i.e. organ)** |  | |  | | | |  | |
| **Total no. of blocks** |  | | | | | | | |
| **Number of sections**  **Specification per block** | No. of blocks | | |  |  | |  |  |
| Section thickness (µM) | | |  |  | |  |  |
| No of sections/block | | |  |  | |  |  |
| **Method** | RNAse-free cutting | serial sections on glass | | | | sections in cups | | |
| **Planned analysis** | Histochem | IHC or (F)ISH | | | | D/ RNA-isolation | | |
| Other, i.e. ………. |  | | | |  | | |
| **Other remarks** |  | | | | | | | |

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| **Histochemical stainings** | | | | |
| **Staining** | HE | PAP | PAS… | other: … |
| **Number of slides** |  |  |  |  |

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| **Immunohistochemical stainings** | | | | | | |
| *IHC staining on Omnis (Dako/Agilent) or BenchMark Ultra (Ventana/Roche)*  *Only for human FFPE samples* | | | | | | |
| **Protocol** | UZA PA\*: see [list IHC UZA pathology department](https://labogidspatana.uza.be/sites/default/files/media/files/2022-07/PAW-009%20L01%20v31.PDF)  optimisation/validation in consultation with researcher  protocol (for Omnis/Benchmark) supplied by researcher | | | | | |
| **Antibody** | # | AB provided by researcher\* | | DAB | Alk Fos or Magenta | |
|  |  |  | |  |  | |
|  |  |  | |  |  | |
|  |  |  | |  |  | |
|  |  |  | |  |  | |
| **Control tissue** | each run | | each slide | | | not required (e.g. internal control present) |
| **Other remarks** |  | | | | | |
| IHC technical quality will be evaluated by pathology department | | | | | | |

\* if protocol of UZA PA is used, then AB is provided by APARTS

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| **In Situ Hybridisaition (ISH)** | | | |
| **Material** | FFPE tissue slides | Cytospins | Other, i.e…… |
| **(F)ISH** | List of probes\*: see [list FISH UZA pathology department](https://labogidspatana.uza.be/sites/default/files/media/files/2022-04/Overzicht%20FISH%20testen%20v13.PDF) | | |
| Probe | # | Probe provided by researcher |
|  |  |  |
|  |  |  |
|  |  |  |
| **Type of analysis** | AMPLIFICATION  BREAK  FUSION  DELETION | | |
| **Number of nuclei for evaluation** | 20 | 50 | Other: ……… |
| **Representative microscopic image needed** | YES  NO | | |
| **CISH** | EBER | Other: … | |

\* if protocol of UZA PA is used, then probe is provided by APARTS

**2 Histopathologic evaluation**

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| --- | --- | --- | --- | --- | --- |
| **Microscopic evaluation** | | | | | |
| **H&E** | evaluate if representative tissue is present | | | | |
| % relevant cells | | | ROI selection and annotation | |
| other: | | |  | |
| **IHC** | # | Scoring system | | | |
| H-score: intensity & % | other scoring system: | | descriptive |
|  |  |  | |  |
|  |  |  | |  |
| **Evaluation by a specific pathologist requested?** | No  Yes – Name: … | | | | |
| **Other remarks** |  | | | | |

**3 Digital Microscopy** (WSI, Ultra-fast scanner, Philips, 40x)

Slides need to be clean without excess mounting media on the (edges of the) cover glass, mounting medium has to be dry and cover glass should NOT stick out. No marking on the slide allowed.

**Please provide an external hard drive for image storage** (0.5-1.5 Gb/image at 40x, depending on tissue area).

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| **Scanning** | |
| **Number of slides** |  |
| **Image format** | Tiff  ISyntax |
| **Required magnification:** | 40x  20x  10x  5x |
| **Back-up storage for 1 month by APARTS** | YES  NO |

**4 Molecular analysis**

|  |  |  |  |
| --- | --- | --- | --- |
| NA extraction | | | |
| **D/RNA isolation of FFPE sections** | # of cases: …. | Macrodissection required?  YES  NO | |
| # of sections /case: ….. | | |
| **cfDNA isolation** | Material:  plasma  urine  CSF  other: ….. | | |
| # of cases: …. | ml/case: …. | Elution volume: …. |
| **Measurement of concentration** | Nanodrop | Qubit DNA BR | Qubit DNA HS |
| Qubit RNA BR | Qubit RNA HS |  |
| **Other remarks** |  | | |

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| PCR | | | |
| **Idylla** | BRAF V600 | NRAS+BRAF | KRAS |
| NSCLC fusion | MSI |  |
| ctEGFR | ctKRAS |  |
| Other: …. | | |
| **Real-time PCR** | HPV Xpert (cyto) | HPV sacace (cyto + FFPE) |  |
| Other: …. | | |
| **Digital droplet PCR** | KRAS G12/13 | KRAS G12C | BRAF V600 |
| EGFR 5 hotspots | EGFR T790M | MYD88 L265P |
| Other: … | | |

Full range of available assays on Idylla platform and QX200 ddPCR platform: [biocartis website](https://www.biocartis.com/en/meet-idylla/idylla-platform) en [bio-rad website](https://www.bio-rad.com/digital-assays).

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| Next Generation Sequencing (NGS) |
| Oncomine Focus Assay (OFA)– DNA panel (SNV and small indels, 23 genes) |
| Oncomine Focus Assay (OFA)– RNA panel for fusions (22 genes) + METex14 skipping & EGFR  VIII |
| Oncomine Comprehensive Assay (OCA) – DNA panel (143 genes) |
| Oncomine Comprehensive Assay (OCA) – RNA panel (22 fusions) |
| BRCA1/2 with Oncomine Research Assay (SNV and small indels) |
| Gynaecology targeted NGS panel (22 genes) |
| Glioma targeted NGS panel including TP53, IDH1/2, 1p19q deletion and EGFR amplification |
| Thyroid targeted NGS panel including POLE (15 genes) |
| TMB with Oncomine Tumor Mutation Load assay |
| CNA by sWGS |
| Oncomine PAN-CANCER cell-free assay (52 genes) |
| Other: …. |

Detailed NGS panel information is available here: [labogids UZA pathologie - NGS](https://labogidspatana.uza.be/analyses/next-generation-sequencing-mutatie-en-fusieanalyse-ffpe).

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| Methylation |
| MGMT promoter hypermethylation by RT-PCR |
| MLH1 promoter hypermethylation by RT-PCR |
| whole genome methylation by Infinium EPIC microarray |
| Other: …. |